A Novel Color Former Organogel for Radiation Detection Systems

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A novel leuco phenoxazine color former containing L-phenylalanine at the protecting group was prepared for radiation detection systems. It forms elongated self-assembled fibers in common organic solvents, leading to that efficient gelation. FE-SEM and FT-IR of the xerogel showed the color former formed a three-dimensional net-like structure. The color former organogel showed sufficient sensitivity to γ rays that it can be used for imaging in heavy particle radiotherapy and has good potential for three-dimensional radiation detection systems.

Radiation has been widely utilized in many fields such as medicine, agriculture, and industry. For example, medical apparatus are sterilized by γ rays without producing hazardous residue. γ Rays are high-energy electromagnetic radiation and the penetration power is very large. As human beings have no ability of directly seeing γ rays, the recognition of γ rays is important by some other method. We have studied the development of the color formers to recognize γ rays easily by visualization.

Recently, many low-molecular-weight organic compounds have been found to be organogelators that can gel various organic solvents. Organogels consist of long nanofibers that are self-assembled as a result of the usual array of noncovalent forces such as hydrogen bonding, van der Waals, and π - π stacking. Noncovalent cross-links among the nanofibers and/or mechanical entanglements create a three-dimensional network. Recent research shows that L-amino acid derivatives had organogelation abilities for most organic solvents. 3

If organogelation ability is introduced to the color former, the diffusion of the generated dyes could be prevented by intermolecular interaction in the three-dimensional color former networks. The gel could be applied in a heavy particle detector and used for imaging in therapeutic instruments.

Here we report the properties of new compound **1** (Scheme 1) in which the phenoxazine moiety is attached to an L-phenylalanine-based scaffold⁴ via a dodecamethylene chain (7.9% yield). The synthesis of **1** was straightforward and was observed by ¹H NMR, ¹³C NMR, FAB-MS, and FT-IR spectra.⁵

Table 1. Gelation properties of color former **1** for common organic solvent^a

Solvent	
Hexane	I
Toluene	S
CHCl ₃	S
CH_2Cl_2	G
CCl_4	G
Ethyl acetate	G
CH_3CN	PG
DMF	S

^aGelation tests were conducted at $80 \,\mathrm{g} \,\mathrm{L}^{-1}$. The following abbreviations are used: P, precipitate; I, insoluble; S, solution; PG, partially gel; and G, gel.

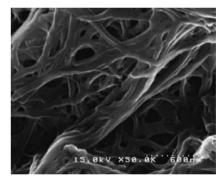
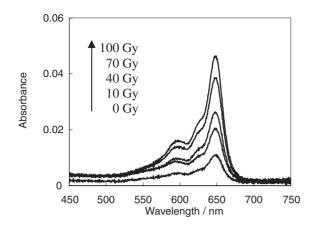


Figure 1. FE-SEM image of xerogel of 1.

Gelation tests were carried out using common solvents (Table 1). Color former **1** was observed to be insoluble in non-polar and protic polar solvents, while it was soluble in several less polar solvents such as ethyl acetate, CHCl₃, and CH₂Cl₂. On estimating the critical gelation concentration (CGC)⁶ of the solvents, the lowest CGC value was obtained: $80 \, \mathrm{g \, L^{-1}}$ when CH₂Cl₂ was used. As the color former gel was easily colored by heating, CH₂Cl₂ was used in the following experiment because of its low boiling point.

The morphological analysis of dichloromethane xerogel of 1 obtained from the dichloromethane gels was studied by field-emission scanning electron microscopy (FE-SEM)⁷ (Figure 1). This analysis showed that 1 was composed of fibrillar self-assembled nanostructures. In order to evaluate the driving forces for gelation, the FT-IR spectra were measured.⁸ The FT-IR spectra of 1 was measured in dichloromethane xerogel. The IR spectra of xerogel of 1 showed bands at 3296 cm⁻¹ (amide ν N-H), 1640 cm⁻¹ (amide ν C=O), and 1546 cm⁻¹ (amide δ N-H), characteristic of hydrogen-bonded amide groups. Such IR shifts indi-



$$\begin{array}{c} \text{Et}_2\text{N} & \xrightarrow{\text{N}} & \text{NEt}_2 \\ \text{N} & \xrightarrow{\text{N}} & \text{Et}_2\text{N} & \xrightarrow{\text{N}} & \text{NEt}_2 \\ & & & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & \\ & & &$$

Figure 2. Spectral changes of **1** to give colored form **2** in dry dichloromethane ([**1**]₀ = 0.25 mM) at 4 h after γ irradiation to the dichloromethane gel of **1**.

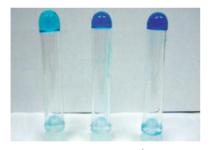


Figure 3. Images of $1([1]_0 = 80 \,\mathrm{g} \,\mathrm{L}^{-1})$ to give colored form **2** after γ irradiation (from left; 0, 40, and 70 Gy).

cate the presence of intermolecular hydrogen-bonding interaction between the amide groups and the carboxyl groups.

UV–vis spectra of the dichloromethane gel of 1 after γ irradiation with ^{60}Co were measured with a solution (0.25 mM) prepared by dilution of the dichloromethane gel of 1 ([1]₀ = $80\,\text{g\,L}^{-1}$) with dichloromethane (Figure 2). The color former gel was slightly colored in the course of preparation. The maximum absorption wavelength increase in the spectra of 1 after γ irradiation coincided with that of $2\cdot\text{Cl}^-$ (649 nm) to indicate that 1 changed to the dye form 2 under γ irradiation. The color change is easily detectable by the naked eye at less than 40 Gy of irradiation (Figure 3). The color change after γ irradiation of

40 Gy suggests that the gel will be useful in heavy particle radiotherapy with heavy particle beams of 40–70 Gy.

In conclusion, a novel leuco phenoxazine color former 1 was prepared and showed efficient gelation in common organic solvents. The gel structure was determined by FE-SEM and FT-IR. This dichloromethane gel of color former 1 showed sufficient color change that γ irradiation of 40 Gy can be detected by the naked eye. A novel color former organogelator will be applied in heavy particle detectors for three-dimensional color former networks in organogel.

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- FAB-MS m/z 853 (M + 1, C₅₂H₈₀N₆O₄); IR (NaCl, cm⁻¹) 3296, 1640, 1546, 1510, 1280, 1241, 740, 701; ¹H NMR (200 MHz, CDCl₃): δ 7.40–7.05 (m, 9H, ArH), 6.51–6.30 (s, 1H, CONH), 6.34-6.13 (s, 2H, ArH), 5.44 (br s, 1H, CONH), 4.70–4.44 (m, 1H, CH₂NH, J = 7.8 Hz), 3.46– 2.88 (m, 24H, NCH₂CH₃, NHCH₂), 2.40–2.05 (s, 2H, NHCOCH₂), 1.83-1.56 (q, 12H, NCH₂CH₃), 1.56-1.13 (m, 20H, CH₂), 1.13-0.80 (t, 3H, CH₃); ${}^{13}C\{{}^{1}H\}$ NMR (126 MHz, (CD₃)₂CO): δ 9.94 (s, CH₃), 11.56 (s, CH₃), 20.43 (s, CH₂), 23.56 (s, CH₂), 24.58 (s, CH₂), 24.70 (s, CH₂), 26.69-27.71 (m, CH₂), 29.71 (s, CH₂), 33.70 (s, CH₂), 36.03 (s, CH₂), 37.03 (s, CH₂), 38.53 (s, CH₂), 42.17 (s, CH₂), 52.49 (s, CH), 97.54 (s, CH), 104.73 (s, CH), 116.23 (s, CH), 123.37 (s, CH), 124.37 (s, CH), 126.15 (s, CH), 127.32 (s, CH), 135.65 (s, CH), 144.47 (m, CH), 154.11 (s, CO), 169.55 (s, CO), 171.38 (m, CO).
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